

Use of acoustic telemetry to evaluate fish movement, habitat use, and protection effectiveness of a coral reef no-take zone (NTZ) in Brazil

Daniel Lino Lippi^{1,*}, Mariana Sofia Coxey², Jay R. Rooker^{3,4}, Sérgio Magalhães Rezende⁵, Michael A. Dance⁶, Ana Lídia Bertoldi Gaspar¹, Mauro Maida¹, Beatrice Padovani Ferreira¹

¹Department of Oceanography, Federal University of Pernambuco, 50740-550 Recife, Brazil
 ²MARE – Marine and Environmental Sciences Centre, ISPA-Instituto Universitário, 1149-041 Lisbon, Portugal
 ³Department of Marine Biology, Texas A&M University at Galveston, Galveston, TX 77553, USA
 ⁴Department of Ecology and Conservation Biology, Texas A&M University, College Station, TX 77843, USA
 ⁵National Center for Research and Conservation of Marine Biodiversity of the Northeast (CEPENE), 55578-000 Tamandaré, Brazil
 ⁶Department of Oceanography and Coastal Sciences, Louisiana State University, Baton Rouge, LA 70803, USA

ABSTRACT: Movement is a key factor that shapes the distribution and structure of fish populations and influences the extent of the benefits provided by conservation and management measures, such as the implementation of marine no-take zones (NTZs). We used visual surveys and acoustic telemetry to investigate density and movement of 2 Brazilian endemic and highly targeted reef fish species inside and outside a coral reef NTZ, and subsequently inferred the effectiveness of the NTZ for protecting these species. To do so, visual surveys were performed on protected and unprotected reefs between 2016 and 2017. Moreover, 20 gray parrotfish Sparisoma axillare and 9 Brazilian snapper Lutjanus alexandrei were tagged with acoustic transmitters and passively monitored from December 2016 to October 2017. For both species, fish densities were significantly higher within the NTZ. Also, both species presented a high residence index over the short term, indicating they were full-time residents of the monitored area until detections were permanently lost. The absence of detections may indicate relocation to deeper reefs, predation, or fishing mortality when fish left the NTZ. Home ranges were small (0.10 to 0.45 km²), and both species presented spatially segregated subgroups within the populations. On average, the percentage of the home ranges within the NTZ was 88% for S. axillare and 95% for L. alexandrei. The results showed that small NTZs that are important to part of the life cycle of a target species are an effective measure to conserve reef fish populations, and also highlight the importance of fisheries management outside NTZs.

KEY WORDS: Acoustic monitoring · Marine protected area · *Sparisoma axillare · Lutjanus alexandrei* · Parrotfish · Snapper · Home range · Spatial ecology · Reserve effect

- Resale or republication not permitted without written consent of the publisher -

1. INTRODUCTION

The creation of marine no-take zones (NTZs) has been considered one of the most effective tools in restoring and preserving biodiversity in coral reef environments (Halpern & Warner 2003, Lester et al. 2009, Edgar et al. 2014). Since the establishment of the first NTZs on coral reefs, a large body of evidence has shown that these protected areas are effective in increasing fish size, abundance, and biomass (Polunin & Roberts 1993, Gell & Roberts 2003, Halpern 2003), recovering populations of endangered species (Afonso et al. 2011, Anderson et al. 2014), restoring the complexity of ecosystems through reestablishment of trophic cascades (Harborne et al. 2008, Leleu et al. 2012), and promoting an overall increase in resilience and complexity within their boundaries (Micheli et al. 2012, Barnett & Baskett 2015).

Effectiveness of NTZs for rebuilding and conserving fish populations is inherently linked to the movement of fishes that they are intended to safeguard (Green et al. 2015). Reef fishes have complex social systems that evolve during their life cycles, connecting habitats and ecosystems through movements and migrations (Jones et al. 2010, Kimirei et al. 2013). Fish movements and resulting changes in habitat use often occur at different spatial and temporal scales, and these dispersive behaviors are influenced by a variety of biological and ecological processes including ontogeny (Dahlgren & Eggleston 2000), reproduction (Afonso et al. 2008, Rhodes et al. 2012), competition (Abesamis & Russ 2005), predation (Bosiger & McCormick 2014, Rooker et al. 2018), and resource availability (Meyer et al. 2000).

Because habitat use and movement may vary considerably among species, the benefits of marine protected areas (MPAs) are species specific, and understanding the timing and degree of movements displayed by species of interest is needed to predict the value of these conservation measures (Green et al. 2015). If NTZs are relatively small in area or unrepresentative of essential habitats, regular movements of species may not be totally encompassed, and individuals will be constantly exposed to a potential risk of fishing mortality, thus compromising the ability of the protected area to promote fish growth and survival within its limits (Kramer & Chapman 1999, Halpern & Warner 2003). On the other hand, one of the main benefits expected from NTZs is their ability to enhance fishery yields and biodiversity outside their boundaries through spillover (Gell & Roberts 2003, Di Lorenzo et al. 2016). In this context, knowledge on short- and long-term movement and habitat-utilization patterns of fish is a key element to the design, management, and evaluation of NTZs in coral reef environments.

Passive acoustic telemetry allows the continuous and simultaneous tracking of individuals within a monitored area over different time scales (Topping et al. 2005, Heupel et al. 2006), and the approach has been commonly used to assess the effectiveness of NTZs (Lea et al. 2016, Di Franco et al. 2018, Lippi et al. 2021) by evaluating different aspects of fish movement ecology including site fidelity (Abecasis & Erzini 2008, Meyer et al. 2010, Harasti et al. 2015), residency (March et al. 2011, Abecasis et al. 2013), home range size (Topping & Szedlmayer 2011, Villegas-Ríos et al. 2013, Garcia et al. 2014), habitat preference (Alós et al. 2011, Marshell et al. 2011, Rooker et al. 2018), mortality events (Khan et al. 2016, Tickler et al. 2019), and reproductive and ontogenetic migrations (Rhodes et al. 2012, Huijbers et al. 2015, Nanami et al. 2018).

In the present study, we used visual surveys and passive acoustic telemetry to characterize the habitat use and movement of 2 endemic and highly targeted fish species within a coral reef NTZ in Brazil: gray parrotfish *Sparisoma axillare* (Steindachner, 1878) (Labridae: Scarini) and Brazilian snapper *Lutjanus alexandrei* Moura & Lindeman, 2007 (Lutjanidae). The primary goals of this study were to estimate residency, movement patterns, and home range of *S. axillare* and *L. alexandrei* to assess the effectiveness of the current NTZ for protecting these species.

2. MATERIALS AND METHODS

2.1. Study area

The monitored area was located on the Tamandaré coral reef complex off the coast of Pernambuco state, in northeastern Brazil (8° 44' S, 35° 6' W), and included the main reef formations inside and around a 2.7 km² well-enforced NTZ established in 1999 (Fig. 1). The NTZ is situated within the larger multiple-use Coral Coast MPA (established in 1997) and is also the core of the marine area of the Municipal Natural Park of Tamandaré Fort (established in 2003). About 30% of the NTZ area is covered by coral reef formations, which are mostly located on its north side and are contiguous to the reefs outside the NTZ.

Four distinct reef areas comprise the seascape within the NTZ: (1) scattered patch reefs, (2) aggregated patch reefs, (3) single-unit reef, and (4) barrier reef. Scattered patch reefs are surrounded by broad sand areas at depths of $\sim 1-3$ m, while aggregated patch reefs are clustered and separated by narrow sand corridors distributed over a deeper area (up to 8 m) with reefs shallower towards the north and outside the NTZ. In both scattered and aggregated patch reef areas, reefs are also surrounded by seagrass and algae beds. Single-unit reef is characterized by a high complexity and relatively isolated structure, with depth ranging from 4 to 12 m. Reef top is exposed during low tide and overhanging



Fig. 1. Study area in the Tamandaré coral reef complex off the northeast coast of Brazil. (A) Locations of the Coral Coast marine protected area (MPA), the no-take zone, and sites where the fish surveys were conducted in the different reef zones: South zone (SZ), No-take zones 1 and 2 (NTZ1, NTZ2), North zones 1 and 2 (NZ1, NZ2). (B) Locations of the acoustic receivers and 50% detection range (150 m). Panels A and B also highlight the different reef areas where fish were collected/released (SUR: single-unit reef; BR: barrier reef; SPR: scattered patch reefs; APR: aggregated patch reefs). (C) Locations of seagrass beds and algae patches in the study area

ledges shade the systems of interconnected caves. Barrier reef consists of an elongated reef formation subjected to stronger wave action. Depth rages from 3 to 6 m. Eastward, beyond the last line of receivers, depth increases continuously up to 8 m.

2.2. Study species and fisheries

Sparisoma axillare has a widespread distribution along the Brazilian coast, from Maranhão (northeastern Brazil) to Santa Catarina (southern Brazil), also including the oceanic islands (Moura et al. 2001). The species is commonly found from shallow coastal reefs and seagrass beds to deeper reefs at depths of up to 54 m (Moura et al. 2001, Feitoza et al. 2005). Current data of landings from the artisanal fisheries in the study area show that *S. axillare* is the most representative species in spear and hook-and-line fisheries in the costal reefs of Tamandaré (Silveira 2018). The species is also an important target for the bottom trap fisheries along the Pernambuco continental shelf (Ribeiro 2004).

Lutianus alexandrei is restricted to northeastern Brazil (from Maranhão to Bahia) and is absent from oceanic islands (Moura & Lindeman 2007). The species undergoes ontogenetic migrations from estuarine/mangrove areas to reefs at depths greater than 50 m (Moura & Lindeman 2007, Aschenbrenner et al. 2016a,b). It is worth noting that the species has only recently been described (Moura & Lindeman 2007), and records of the species had been misidentified as L. jocu and/or as the Caribbean species L. griseus and L. apodus up to that date (Moura & Lindeman 2007). Brazil is lacking official landing statistics since 2008 (Freire et al. 2021), and therefore such misidentifications are incorporated in the available records for the Lutjanidae fisheries. L. alexandrei is captured by different gears (line, gillnet, and bottom traps) from both small-scale and commercial reef fisheries of northeastern Brazil, where it is part of the Lutjanidae complex (Ribeiro 2004, Frédou et al. 2006).

2.3. Reef fish surveys

Visual surveys using belt transects of 100 m² (5 m wide by 20 m long) were used to compare fish density and size composition for the 2 target species. During surveys, S. axillare and L. alexandrei individuals were identified and counted, and total length (TL, cm) was estimated and recorded by trained observers. A minimum of 4 replicate transects were surveyed at each site during low tides, with no sampling taking place when underwater visibility was lower than 4 m. Surveyed sites were randomly chosen within 5 different reef areas, always along reef crests. A total of 314 transects were surveyed during 2 consecutive dry seasons (January to April 2016 and September 2016 to March 2017). Visual surveys conducted within the NTZ corresponded to the single-unit reef area (NTZ1 = 98 transects) and patch-reef area (protected portions of scattered patch reefs and aggregated patch-reef areas, NTZ2 = 32 transects). Surveys outside the NTZ were performed on the reefs located beyond the southern (SZ = 48 transects) and northern (NZ1 = 74 and NZ2 =62 transects) borders of the NTZ (Fig. 1).

2.4. Acoustic monitoring system

From December 2016 to August 2018, an array of 17 omnidirectional acoustic receivers (VR2W-69kHz, VEMCO) were used to monitor the movements and habitat use by tagged *S. axillare* and *L. alexandrei* individuals (Fig. 1). Preliminary range tests conducted inside the NTZ demonstrated that, in addition to distance between receivers and transmitters, tide level also had a negative effect on detection probability. A minimum detection rate of 50% was obtained for transmitters at 150 m distance from the receiver during preliminary trials, and therefore this range was used as a guideline to design the array. At 300 m, average detection probability dropped to below 5% during high tide hours (V. M. Giacalone et al. preprint, doi:10.13140/RG.2.2.35735.57768).

In most cases, receiver positioning allowed for overlapping detection ranges, and the total area covered by the array was approximately 1 km², including more than 80% of the reef area within the NTZ and the reef formations adjacent to its northern border. To identify fish movement to unprotected areas, 3 receivers were placed outside the NTZ, on the reef continuum beyond its northern boundary. Depth inside the monitored area ranged from 1 to 12 m.

As reef tops may be exposed (or just below the surface) during low tide, receivers were placed over the sandy bottom around the reefs, moored in a PVC pipe attached to a concrete base to ensure vertical positioning and avoid dragging. Data downloads, battery checks, and clearing of fouling organisms took place every 1–2 mo. One receiver located at the southernmost part of the barrier reef (Fig. 1) was lost in the 2017 winter due to severe sea conditions.

2.5. Fish collection and tagging

From December 2016 to May 2017, 20 *S. axillare* and 9 *L. alexandrei* were caught and tagged. All fish were caught inside the NTZ (Fig. 1). To reduce possible school influence on parrotfish behavior (Welsh & Bellwood 2012a) and better represent the local population, collections of *S. axillare* were equally distributed among the 4 reef areas: single-unit reef (n = 5), scattered patch reefs (n = 5), aggregated patch reefs (n = 5), and barrier reef (n = 5). For *L. alexandrei*, fish were captured in the single-unit reef (n = 5), scattered patch reefs (n = 3), and aggregated patch reefs (n = 1).

Individuals were caught with hand nets by SCUBA divers at night when fish were either 'sleeping' (S. axillare) and/or with reduced flight capability (L. alexandrei). This way, impact was minimized, as only individuals from the target species and with a desirable body size (S. axillare >24 cm TL and L. alexandrei >19 cm TL) were caught. This collection method also reduced the disturbance to the local fish community and the risk of injury caused by fishing gears. Individuals were transported to a research facility on land and kept in 1000 l holding tanks with direct seawater and monitored for 24 h prior to tag implantation. S. axillare were implanted with individually coded V9-4L acoustic transmitters (30 to 90 s delay, 476 d expected lifetime, 4.5 g tag weight, VEMCO) while L. alexandrei were implanted with V8-4L acoustic transmitters (30 to 90 s delay, 150 d expected lifetime, 2.1 g tag weight, VEMCO). Prior to surgery, fish were transferred to a smaller tank and anesthetized in a 0.04 ml l⁻¹ seawater and eugenol solution (Pastor et al. 2009, Lee et al. 2015, Honda et al. 2016, 2017), weighed (total weight, nearest g), and measured (TL, nearest mm). Individuals were then placed in a 'V' shaped bed, and scales were plucked from the ventral midline to expose the tag-insertion area. Transmitters were cleaned in povidone-iodine to prevent infection (Abecasis et al. 2009, Ferguson et al. 2013) and then surgically inserted into the peritoneal cavity through a 1.5-2.0 cm incision, which was closed with 2 stitches of absorbable 4/0 monofilament suture. Direct seawater flow was maintained

through the gills by a water pump allowing fish to breathe normally throughout the procedure. After surgery, individuals were returned to the holding tanks and observed for a 24 h period prior to release. Releases occurred the following morning, at the capture site of each individual. All tagged *S. axillare* were initial-phase females, with a mean (\pm SD) size of 26.5 \pm 2.3 cm TL (24.2–33 cm) and weight of 296.3 \pm 73.5 g (Table 1). Sex of *L. alexandrei* was not determined, and mean size and weight were 22.5 \pm 2.3 cm TL (19.5 to 25.2 cm) and 186.3 \pm 60.3 g (Table 2). For all individuals, the tag-to-body weight relationship

never exceeded 2% ($\overline{x} = 1.49 \pm 0.14\%$ CI, range = 0.83–1.95%) of fish total weight in air, as recommended by the manufacturer of the transmitters.

2.6. Data analysis

Fish density and TL were compared among reef sites using the non-parametric Kruskal-Wallis and post hoc Dunn's pairwise ranked tests. For the purpose of fish density analyses, *S. axillare* were divided in 2 size categories: ≤ 20 and ≥ 20 cm, as followed by the Reef Check protocol (Ferreira et al. 2018); *L. alexandrei* had a single size category.

For the acoustic telemetry data, prior to statistical analysis, spurious detections (i.e. any detection from a single fish occurring alone within a 24 h period) were removed to prevent any false-positive detections (March et al. 2011, Harasti et al. 2015). Detections obtained within the first 24 h post-release were also excluded from all individuals to avoid potentially negative effects of the tagging procedure on fish behavior (Honda et al. 2016). To identify groups of individuals with distinct spatial occupation patterns within each species, the relative number of detections on each receiver was calculated for each fish. Proportions were then arcsine-square root transformed (Meyer et al. 2010, Topping & Szedlmayer 2011) and a similarity matrix among fish was built for each species based on the Bray-Curtis similarity index. Statistically distinct spatial groups were then identified through hierarchical cluster analysis followed by a SIMPROF procedure (Clarke & Gorley 2006). Differences in TL were tested among spatial groups for each species using ANOVA and Student's t-test.

Permanence within the monitored area was quantified for each fish by 2 indices (Abecasis et al. 2013, 2015, Özgül et al. 2019): (1) a residency index expressed as the number of days an individual was actually detected (D_D) divided by the detection period of the individual (number of days between release and the last detection, D_P) and (2) a weighted residency index calculated as:

Weighted residency index =

$$(D_D/D_{max}) \times (D_P/D_{max})$$
(1)

where D_{max} is the maximum number of possible monitoring days for each species based on estimate battery life (*S. axillare* V9 lifetime = 476 d; *L. alexandrei* V8 lifetime = 150 d). The end of study date was considered as D_{max} when the removal of the receiver array occurred before the expected lifetime of the transmitter was over. For both indices, values range from 0 (no residency) to 1 (permanent residency).

To estimate the extent of space used by each individual, centers of activity (COAs) were first calculated for each fish at 60 min intervals (Simpfendorfer et al. 2002). This method uses the weighted means of the number of detections registered by each receiver to provide estimated hourly fish positions (mean latitude and longitude). The nonparametric kernel utilization distribution (KUD) was then used to calculate home range (defined as 95% KUD) and core range (defined as 50 % KUD) areas, based on the COA estimates over the entire detection period of the individual. KUDs were estimated based on bivariate fixed kernels (Worton 1989) over a 25×25 m cell grid and a fixed bandwidth smoothing factor (h) of 150 m (corresponding to the 50% detection probability range). As observed during diurnal and nocturnal dives in the monitored reefs, both species are highly reef attached and the use of bare sand/mud areas is limited to the immediate reef vicinity. Therefore, the entire 95 and 50% KUD areas were considered an overestimation of the actual space used by both species. To produce more reliable home range and core range estimates, the surrounding sand-covered areas located more than 50 m away from any reef formation were excluded from the initially calculated 95 and 50% KUDs. The selection of an area of interest reduced the home range and core range areas of S. axillare by 25.1 \pm 10.6 % CI and 21.2 \pm 10.4 % CI, respectively. For L. alexandrei, home range and core range areas were reduced by $34.0 \pm 13.6\%$ CI and $24.9 \pm 16.3\%$ CI, respectively. The majority of seagrass and macroalgae patches present in the monitored area were included within the 50 m limit range. Finally, individual home range and core range areas were overlapped with the NTZ boundary to assess the percentage of home range and core range areas located inside the NTZ.

To investigate space-use sharing between the different spatial groups within species, mean home range

ig data for tagged Sparisoma axillare (n = 20) in the coastal reefs of the Tamandaré coral reef complex. Total length (TL, cm), cap-	ups (SPR: scattered patch reefs, APR: aggregated patch reefs, BR: barrier reef, SUR: single-unit reef), detection period (D _p), days	index (I_{WR}) , residency index (I_R) , number of centers of activity (COAs), percentage of home range located inside the no-take zone	nd percentage of core range located inside the NTZ (%CR _{in}). Dashes (–) indicate no detection; NA: not available
Table 1. Summary of the monitoring data for tagged Sparisoma a	ture/release areas and spatial groups (SPR: scattered patch reefs	detected (D_D) , weighted residency index (I_{WR}) , residency index $(I_f$	(NTZ) (%HR _{in}), and percentage of core range lo

Fish ID	TL (cm)	Date released (d/mo/yr)	Capture/ release areas	Spatial groups	No. of detections	$D_{ m P}$	$D_{ m D}$	$I_{ m WR}$	All O	– I _R ——— utside NTZ	COAs	Home I All	range (km²) % HR _{in}	Core n All	ange (km²) % CR _{in}
SPAAXI #01	26.2	13/12/2016	SUR	SUR	13 002	51	51	0.01	1.00	I	837	0.10	100	0.02	100
SPAAXI #02	a 26.1	13/12/2016	SUR	NA	87	01	01	NA	NA	NA	NA	NA	NA	NA	NA
SPAAXI #03	25.5	12/01/2017	SUR	SUR	4091	18	18	0.001	1.00	I	314	0.14	100	0.04	100
SPAAXI #04	25.0	12/01/2017	SPR	NA	1252	25	25	0.003	1.00	I	212	0.44	98.9	0.11	100
SPAAXI #05	a 31.5	14/01/2017	SUR	NA	34234	480	325	NA	NA	NA	NA	NA	NA	NA	NA
SPAAXI #06	25.0	14/01/2017	SPR	SPR	733	23	23	0.002	1.00	0.13	111	0.34	86.8	0.07	99.7
SPAAXI #07	27.7	14/01/2017	SPR	SPR	942	17	17	0.001	1.00	0.18	117	0.42	85.1	0.07	97.2
SPAAXI #08	33.0	20/01/2017	BR	BR	437	107	62	0.03	0.58	0.07	158	0.40	80.8	0.07	100
SPAAXI #09	27.0	20/01/2017	BR	BR	63	26	20	0.002	0.77	I	37	0.25	100	0.07	100
SPAAXI #10	25.4	20/01/2017	BR	BR	490	55	50	0.01	0.91	I	217	0.18	100	0.05	100
SPAAXI #11	27.5	20/01/2017	BR	BR	125	91	39	0.02	0.43	I	65	0.16	100	0.04	100
SPAAXI #12	a 25.0	20/01/2017	BR	NA	19	00	04	NA	NA	NA	NA	NA	NA	NA	NA
SPAAXI #13	25.2	26/01/2017	APR	APR	180	56	39	0.01	0.70	I	86	0.28	82.9	0.07	100
SPAAXI #14	a 29.0	26/01/2017	APR	NA	484	03	02	NA	NA	NA	NA	NA	NA	NA	NA
SPAAXI #15	25.0	30/03/2017	SPR	SPR	7190	58	58	0.02	1.00	0.26	840	0.22	72.1	0.05	97.9
SPAAXI #16	24.5	30/03/2017	SPR	SPR	12353	180	34	0.03	0.18	0.01	377	0.21	78.7	0.05	99.9
SPAAXI #17	26.8	30/03/2017	SUR	SUR	13821	38	38	0.01	1.00	I	555	0.19	100	0.06	100
SPAAXI #18	24.3	28/04/2017	APR	APR	3017	159	151	0.14	0.95	0.03	711	0.31	84.1	0.06	100
SPAAXI #19	25.6	28/04/2017	APR	SPR	717	28	25	0.004	0.89	0.61	136	0.45	58.8	0.10	49.0
SPAAXI #20	24.2	28/04/2017	APR	APR	866	187	134	0.15	0.71	0.01	355	0.34	86.6	0.07	100
^a Fish that pr	ematurel	y left the array,	lost the tag, oi	r died											
Table 2. Sumr	nary of th	ie monitoring d	ata for tagged	Lutjanus	i) alexandrei (j	n = 9) i	n the	coastal :	reefs of	the Tamand	aré cora	ıl reef cc	mplex. Abbre	eviations	as in Table 1
Fish ID	TL (cm)	Date released	Capture/	Spatial	No. of	$D_{ m P}$	$D_{ m D}$	$I_{\rm WR}$		I _R	COAs	Home 1	range (km²)	Core r	ange (km ²)

ange (km²) % CR _{in}	100 100	100	100	100	99.2	100	100	100
Core r All	0.05 0.06	0.03	0.08	0.07	0.05	0.05	0.04	0.03
range (km²) % HR _{in}	100 100	100	86.2	100	74.2	100	96.9	100
Home All	$0.15 \\ 0.19$	0.15	0.40	0.26	0.27	0.15	0.19	0.12
COAs	276 1400	1801	1066	113	2139	2345	582	126
— I _R —— Dutside NTZ	1 1	I	I	I	0.74	I	I	I
All C	$1.00 \\ 0.96$	1.00	0.93	0.89	1.00	1.00	0.88	1.00
$I_{ m WR}$	0.03 0.99	0.81	0.67	0.01	0.91	1.00	0.45	0.01
$D_{ m D}$	24 146	135	119	17	143	150	94	16
$D_{ m P}$	24 152	135	127	19	143	150	107	16
No. of detections	$3122 \\ 29810$	14154	3261	1893	39736	45837	2032	885
Spatial groups	SUR SUR	SUR	SPR	SUR	SPR	SUR	SPR	SUR
Capture/ release areas	SUR SUR	SUR	SPR	SPR	SPR	SUR	APR	SUR
Date released (d/mo/yr)	13/12/2016 13/12/2016	12/01/2017	12/01/2017	12/01/2017	14/01/2017	30/03/2017	28/04/2017	05/05/2017
rL (cm)	24.0 21.5	24.5	25.2	23.1	24.5	19.5	20.4	19.5
Fish ID	LUTALE #01 LUTALE #02	LUTALE #03	LUTALE #04	LUTALE #05	LUTALE #06	LUTALE #07	LUTALE #08	LUTALE #09

and mean core range areas were calculated for each spatial group based on the total number of COAs estimated for all fish from the group. Estimates of mean home range and core range followed the same procedures used to calculate individual KUDs. An overlap index (OI) was then calculated for each spatial group as:

$$OI = OV_{(KUD_{i}, KUD_{i+1})} \div (KUD_{i} + KUD_{i+1})$$
(2)

where $OV_{(KUDi, KUDi+1)}$ is the overlap area between the mean home range or core range areas of 2 spatial groups, and $(KUD_i + KUD_{i+1})$ is the combination of the mean home range or core range areas of 2 spatial groups. Values range from 0 (no overlap, groups are spatially segregated) to 1 (mean home range or core range are completely overlapped and groups show no spatial segregation).

KUD analyses were performed using the 'Home Range Tools' extension for ArcGIS (Rodgers et al. 2015). Selection of KUD areas of interest, overlap between home range and core range areas and NTZ boundary, as well as the overlapped and combined mean home range and core range areas were obtained using geoprocessing tools on ArcGIS 10.4. Differences in residency index, weighted residency index, and home range and core range areas between species and among spatial groups were assessed using ANOVA and Student's t-test (or the non-parametric Kruskal-Wallis and Mann-Whitney U-test when parametric assumptions were not met). Also, Spearman's rank correlations were used to investigate if fish TL was significantly correlated with residency index, weighted residency index, home range, and core range. Analyses were conducted in Statistica v. 8.0 (Statsoft) and PRIMER v. 6.1 (Clarke & Gorley 2006). Significance was tested at $\alpha = 0.05$.

2.7. Ethics statement

This research was approved by the Ethics Committee and Animal Use of the Federal University of Pernambuco (CEUA-UFPE Number: 23076.007810/ 2015-01) and Chico Mendes Institute for Biodiversity Conservation (ICMBIO – Sisbio License: 45992).

3. RESULTS

3.1. Reef fish surveys

Fish densities for both species were significantly higher inside the Tamandaré NTZ, with *Sparisoma*

axillare >20 cm forming large roving schools along reef crests and *Lutjanus alexandrei* aggregating in large numbers to rest inside caves. Outliers and extreme outliers indicate the occurrence of those schools, which for *S. axillare* could include up to 70 individuals (Fig. 2). Densities were significantly higher in NTZ1 (corresponding to the single-unit reef area) for *S. axillare* >20 cm (Kruskal-Wallis H = 36.2, p < 0.001) and for *L. alexandrei* (H = 39.8, p < 0.001), and in NTZ2 (corresponding to scattered patch reefs and aggregated patch reefs) for smaller *S. axillare* ≤ 20 cm (H = 37.5, p < 0.001) (Fig. 2). For *S. axillare*, fish TL was also significantly higher in NTZ1 (H =36.2, p < 0.001), whereas larger *L. alexandrei* were observed in NZ2 (H = 16.5, p < 0.01) (Fig. 2).

3.2. Acoustic telemetry

The tagging and monitoring data of the 20 S. axillare and 9 L. alexandrei individuals are summarized in Tables 1 & 2, respectively. Three S. axillare were detected for a short period of ≤6 d (ID nos. SPAAXI #02, #12 and #14). One S. axillare (SPAAXI #05) was assumed to be dead a few days after tagging in a detectable location in the array, as this fish was continuously being detected by only 1 receiver and detection ceased only after the estimated lifespan of the transmitter was over. Therefore, these 4 S. axillare individuals were excluded from all subsequent analyses. Only 1 fish (SPAAXI #11) seemed to have its monitoring discontinued following receiver loss, as this fish was being recorded exclusively by this receiver and ceased to be detected only 3 d before the last date of download. Two other S. axillare individuals were constantly being detected by this receiver while it was still in place; however for both fish, detection ceased more than 40 d prior to receiver loss. The remaining 16 S. axillare and all 9 L. alexandrei were monitored for 16 to 187 d, from December 2016 to October 2017 (Fig. 3A,B).

Even with a shorter transmitter battery lifespan, *L. alexandrei* had a higher number of detections per fish ($\bar{x} = 15637 \pm 18018$ SD) and longer detection period (97 ± 59.5 d) than *S. axillare* (3705 ± 5002 detections and 70 ± 58.3 d), although individual variation within each species was high (Tables 1 & 2). Based on the relative number of detections for each fish at each receiver, hierarchical cluster analysis followed by a SIMPROF procedure revealed 4 significantly distinct clusters for *S. axillare*: barrier reef, scattered patch reefs, aggregate patch reefs, and single-unit reef spatial groups (Fig. 3C), and 2 significantly distinct cluster



Fig. 2. Fish density (left column) and total length (right column) of *Sparisoma axillare* (SPAAXI) and *Lutjanus alexandrei* (LUTALE) obtained by the visual surveys performed in the Tamandaré coral reef complex. For density estimates, *S. axillare* was divided in 2 size categories (>20 and \leq 20 cm total length). Letters above error bars (a–l) indicate the results of Dunn's pairwise ranked tests. Solid line within the box indicates the median, box represents the interquartile range (lower = 25th percentile, upper = 75th percentile), whiskers extend to the largest value within 1.5× the interquartile range, outliers outside this range are represented by black dots, and extreme outliers (values beyond 3× the interquartile range) are represented by asterisks. The top-right panel shows the location of the surveyed site (abbreviations as in Fig. 1)

ters for *L. alexandrei*: scattered patch reefs and singleunit reef spatial groups (Fig. 3D), which corresponded to groups of individuals with distinct spatial occupation patterns. For both species, spatial groups were mostly composed of the individuals captured in the same reef area (corresponding to 94% of *S. axillare* and 78% of *L. alexandrei* individuals). Only 1 *S. axillare* (SPAAXI #19, Fig. 3C) and 2 *L. alexandrei* (ID nos. LUTALE #05 and #08, Fig. 3D) had more detections on reef areas other than their original capture locations. Fish SPAAXI #04 was not included in any spatial group. Neither species showed significant differences in fish TL among spatial groups (*S. axillare*: ANOVA, F = 2.39, p = 0.12; *L. alexandrei*: Student's *t*-test, t = 0.82, p = 0.44).

On average, *S. axillare* individuals were detected on 11 ± 9.7 % SD of the maximum number of monitoring days (lifetime of V9 = 476 d, or end of the study), resulting in low weighted residency index values for the species (0.03 ± 0.05) (Table 1, Fig. 3A). For *L. alexandrei*, weighted residency index values were considerably higher (0.54 ± 0.42) as fish remained in the monitoring area for 62.5 ± 39% of the maximum number of monitoring days (expected lifetime of V8 =



Fig. 3. (A,B) Detection plots of fish daily presence–absence at the monitored area and (C,D) hierarchical cluster analysis followed by a SIMPROF procedure indicating the formation of significantly distinct groups of fish with distinct spatial occupation patterns (red dotted lines) for tagged *Sparisoma axillare* (SPAAXI; A,C) and *Lutjanus alexandrei* (LUTALE; B,D) in the Tamandaré coral reef complex. Original capture/release areas and fish spatial groups formed after clustering with correspondent colors: barrier reef (BR, green), aggregate patch reefs (APR, red), sparse patch reefs (SPR, orange), and single-unit reef (SUR, blue). Fish SPAAXI #04 (gray) was not included in any group. Fish release dates (black circles), days with detections outside the no-take zone (black bars), estimated lifetime of transmitters (465 d for *S. axillare* V9 and 150 d for *L. alexandrei* V8) (asterisks), and removal of receiver array (end of study, vertical red line) are also shown

150 d) (Table 2, Fig. 3B). For both species, no significant differences in weighted residency index were observed among fish groups (*S. axillare*: Kruskal-Wallis, H = 4.04, p = 0.25; *L. alexandrei*: Mann-Whitney, U = 8, p = 0.90). Both species had high residency index values (*S. axillare*: 0.82 ± 0.24 ; *L. alexandrei*: 0.96 ± 0.05) (Tables 1 & 2), with no significant difference between them (Mann-Whitney, U = 52, p = 0.24). For *S. axillare*, higher residency index values were observed for the single-unit reef group (residency index = 1 for all fish), followed by scattered patch reef (0.79 ± 0.14) groups, whereas lower values were obtained for the barrier reef group (0.67 ± 0.21). Significant differences in residency index were ob-

served for *S. axillare* between single-unit reef and barrier reef groups (Mann-Whitney, U = 0, p = 0.02) and between single-unit reef and aggregated patch reef groups (Mann-Whitney, U = 0, p = 0.03). For *L. alexandrei*, no statistical difference was found between the 2 groups (Mann-Whitney, U = 5, p = 0.30). No correlations were found between fish TL and weighted residency index (Spearman rank correlation; *S. axillare*, $r_s = -0.02$, p = 0.93 and *L. alexandrei*, $r_s = -0.01$, p = 0.97) or residency index (Spearman rank correlation; *S. axillare*, $r_s = -0.42$, p = 0.10 and *L. alexandrei*, $r_s = 0.07$, p = 0.85).

The estimated home range (95% KUD) areas ranged from 0.10 to 0.45 km² ($\bar{x} = 0.28 \pm 0.11$ km² SD) for *S. axillare* (Table 1) and from 0.12 to 0.40 km²

 $(0.21 \pm 0.09 \text{ km}^2)$ for *L. alexandrei* (Table 2). Regarding the core range (50% KUD) estimates, areas ranged from 0.02 to 0.11 km² (0.06 ± 0.02 km²) for *S. axillare* (Table 1) and from 0.03 to 0.08 km² (0.05 ± 0.02 km²) for *L. alexandrei* (Table 2). Both home range and core range sizes did not differ significantly between species (Student's *t*-test, *t* = 1.48, p = 0.15 and *t* = 1.31, p = 0.20, respectively) or among spatial groups within each species (Table 3). For *S. axillare*, no correlations were found between fish TL and home range or core range size (Fig. 4A). For *L. alexandrei*, the influence of fish TL on home range and core range sizes was also not significant (Fig. 4B).

Nine individuals were recorded outside the NTZ, but only 3 of them (all from the scattered patch reef group) left the protected area on a regular basis: SPAAXI #15 (26% of monitoring period), SPAAXI #19 (61%), and LUTALE #06 (74%) (Tables 1 & 2, Fig. 3A,B). SPAAXI #19 also had the highest number of detections outside the NTZ (65% of all detections). All S. axillare and L. alexandrei from single-unit reef groups were never detected outside the NTZ. Regarding the percentage of fish home range and core range located inside the NTZ (% HR_{in} and % CR_{in} , respectively), most S. axillare individuals (except for SPAAXI #19) and all L. alexandrei had a % HR_{in} higher than 70% (*S. axillare*: $\bar{x} = 88.4 \pm 6\%$ CI; L. alexandrei: 95.2 \pm 4.5%) and a %CR in over 95% (S. axillare: 96.5 ± 6.2%; L. alexandrei: 99.9 ± 0.1%) (Tables 1 & 2). No significant differences between species were observed for both $\%HR_{in}$ (Mann-Whitney, U = 45, p = 0.16) and %CR_{in} (Mann-Whitney, U = 52, p = 0.24). For *S. axillare*, all fish from the single-unit reef spatial group had their home ranges located entirely inside the NTZ, whereas 1 fish from barrier reef (%HR $_{in}$ = 95.2 ± 9.4 %) and all fish from scattered patch reefs (%HR_{in} = 76.3 ± 9.9 %) and aggregated patch reefs (% $HR_{in} = 84.5 \pm 2.1\%$) had part of their home ranges outside the NTZ (Fig. 5A). Similarly, all L. alexandrei from the singleunit reef group used only areas inside the NTZ, while all fish from scattered patch reefs (%HR_{in} = 85.7 \pm 12.9%) had part of their home ranges beyond the NTZ boundary (Fig. 5B). Significant differences in %HR_{in} were observed among S. axillare spatial groups (Kruskal-Wallis, H = 7.8, p = 0.04), specifically between single-unit reef and scattered patch reefs (Mann-Whitney pairwise, U = 0, p > 0.03). For L. alexandrei, a significant difference in %HR_{in} was also found between single-unit reef and scattered patch reefs groups (Mann-Whitney pairwise, U = 0, p > 0.03). Regarding the %CR_{in}, only 5 *S. axillare* (all from scattered patch reefs group, $%CR_{in}$ = 88.8 ± 19.5%) and 1 L. alexandrei (also from scattered patch reefs group, %HR_{in} = 99.7 ± 0.4 %) had part of their core ranges outside the NTZ.

In general, fish from the different spatial groups used their respective capture/release reef areas more intensively (Fig. 6A,B). Therefore, low overlap index (OI) values were observed among the mean home range areas of the spatial groups for both *S. axillare* ($\bar{x} = 0.14 \pm 0.16$ SD) and *L. alexandrei* (OI = 0.22) (Fig. 6C,D). A moderate overlap (OI = 0.44) was only observed between *S. axillare* from scattered patch reefs and aggregated patch reefs. For both species, fish from the different spatial groups used completely distinct mean core range areas resulting in no overlap (Fig. 6A,B).

4. DISCUSSION

Positive effects of NTZs for fish communities, characterized by increased fish density and larger body size within the protected area, have been well documented worldwide (Halpern 2003, Lester et al. 2009, Malcolm et al. 2018, Gilchrist et al. 2020) and in Brazil (Floeter et al. 2006, Anderson et al. 2014, 2020).

Table 3. Mean ± SD home range and core range areas (km²) for *Sparisoma axillare* and *Lutjanus alexandrei* spatial groups (SUR: single-unit reef; SPR: scattered patch reefs; APR: aggregated patch reefs; BR: barrier reef). Results of ANOVA and Student's *t*-test are shown

Source	SUR	SPR	APR	BR	df	– ANOVA – <i>F</i>	р
Sparisoma axillare							
Home range	0.14 ± 0.04	0.33 ± 0.11	0.31 ± 0.03	0.25 ± 0.11	3	2.92	0.08
Core range	0.04 ± 0.02	0.07 ± 0.02	0.07 ± 0.01	0.05 ± 0.01	3	1.84	0.19
					S	Student's <i>t</i> -tes	t —
Lutjanus alexandrei					df	t	р
Home range	0.17 ± 0.05	0.29 ± 0.10			7	2.30	0.06
Core range	0.05 ± 0.02	0.06 ± 0.02			7	0.91	0.39



Fig. 4. Spearman rank correlation between total length and home range (black circles and font color) and core range (grey circles and font color) sizes for (A) *Sparisoma axillare* and (B) *Lutjanus alexandrei* in the Tamandaré coral reef complex

Our results also revealed a positive effect on fish density and size within the Tamandaré NTZ for both gray parrotfish *Sparisoma axillare* and Brazilian snapper *Lutjanus alexandrei*, corroborating previous studies in the area (Ferreira & Maida 2007). *S. axillare* >20 cm and *L. alexandrei* showed markedly higher densities in the single-unit reef area. In this area, *L. alexandrei* often aggregate to rest in caves

located underneath reef crests, forming large groups. Indeed, this behavior has been previously described to be a characteristic of many reef fishes that seek shelter and school for protection (McClanahan et al. 2007, Beck et al. 2014). Larger *S. axillare* (>20 cm) formed roving schools that could include up to around 70 individuals. This schooling behavioral strategy, efficient to avoid predators and competitors, makes those schools a target to fisheries, and as a consequence, they tend to shrink in numbers in heavily fished areas (Sadovy & Domeier 2005, Hamilton et al. 2016, Guerra et al. 2020). The presence of large schools in the Tamandaré NTZ shows a positive effect of the protection inside the NTZ.

Smaller S. axillare (≤ 20 cm), by contrast, were more abundant in the patch reef areas (scattered patch reefs and aggregated patch reefs) of the NTZ, indicating a preference for this shallower habitat that is continuous inside to outside the NTZ. This area is less complex than the single-unit reef area, but has closely spaced shelter, and is less populated by larger fish, including predators, optimizing safe foraging. In addition, this area includes seagrass and algal beds, which are important as nursery areas for S. axillare (Feitosa & Ferreira 2015, Eggertsen et al. 2017). The observed size distribution of S. axillare indicates that recruitment occurs in the area, and that the patch reef areas are the preferred sites. For L. alexandrei, very few individuals smaller than 15 cm were observed, an expected pattern since the species has been shown to spend its first years in estuarine/ mangrove areas, moving to reefs during their third or fourth year of life (Aschenbrenner et al. 2016a,b).

Despite the low weighted residency index values for S. axillare and some L. alexandrei individuals, nearly all tagged individuals showed a high residency index over the short term, indicating that both species were full-time residents in the monitored area until detections were permanently lost. Comparable detection periods were observed for the Mediterranean parrotfish S. cretense in a similar sized marine reserve (0.83 km²) in the Mediterranean (La Mesa et al. 2012) and for the stoplight parrot S. *viride* and schoolmaster *L. apodus* in a relatively larger (9.56 km²) Caribbean marine reserve (Garcia et al. 2015). However, long-term studies have shown high residency of up to 937 d for S. cretense (Afonso et al. 2008) and up to 363 to 1096 d for other Lutjanus species in natural and artificial habitats (Tin-Han et al. 2014, Huijbers et al. 2015, Williams-Grove & Szedlmayer 2016).

A decline in the number of detected fish throughout the monitoring period is usually observed in



Fig. 5. Mean percentage of home range areas located inside the Tamandaré no-take zone (NTZ) (%HR_{in}) for (A) *Sparisoma axillare* and (B) *Lutjanus alexandrei* spatial groups: single-unit reef (SUR), scattered patch reefs (SPR), aggregated patch reefs (APR), and barrier reef (BR). Error bars indicate 95% confidence interval. Map on the right highlights the location of the different reef areas that correspond to each of the spatial groups

acoustic telemetry studies (TinHan et al. 2014, Garcia et al. 2015, Wolfe & Lowe 2015), which can be mostly explained by natural mortality (e.g. predation), fishing mortality, and movement beyond the detection range of receivers (Khan et al. 2016, Brownscombe et al. 2019). Another possibility is that detection loss of tagged fish may be due to a transmitter malfunction or premature failure of the battery; however, equipment failure was not addressed as a relevant reason for detection loss in studies using similar equipment (Afonso et al. 2016, Khan et al. 2016). Moreover, in our study, transmitters used on 2 L. alexandrei and the one used on SPAAXI #05 were detected throughout their entire estimated battery lifetime. For the 3 S. axillare with noticeably short detection periods (SPAAXI #02, #12, #14) and for SPAAXI #05 (assumed to be dead a few days after release) mortality as a consequence of the tagging procedure should also be considered (Khan et al. 2016, Brownscombe et al. 2019).

Predation mortality is an expected effect of NTZs, as the initial increase in the abundance of prey attracts and even sustains populations of larger predators (Russ & Alcala 1996, Steneck 1998). On the Tamandaré coral reef complex, large predators such as goliath grouper *Epinephelus itajara*, green moray eel *Gymnothorax funebris*, cubera snapper *L. cyanopterus*, and great barracuda *Sphyraena barracuda* are commonly seen inside the NTZ, as are signs of predation attempts on the resident prey fishes (B. P. Ferreira & D. L. Lippi pers. obs.). Seven parrotfish (SPAAXI #02, #03, #07, #13, #14, #16, #17) had their last detections while sheltering at night. Since parrotfishes are strictly diurnal (Hobson 1975), it seems unlikely that these individuals would have left the monitoring area undetected during the night, and thus predation seems to be a reasonable explanation for fish disappearance in the present study.

Illegal fishing can also be a reason for detection loss in acoustic telemetry studies conducted inside NTZs, especially for areas with limited enforcement, such as large and remotely located NTZs (TinHan et al. 2014, Tickler et al. 2019). Although illegal fishing can also be a reason for detection loss in acoustic telemetry studies, fishing mortality inside the Tamandaré NTZ is unlikely as the area is relatively small, with well-defined boundaries and located close to the shore in front of the research facility. Such characteristics make the Tamandaré NTZ a fully enforced area that is easily monitored from both boat and land. However, fishing is allowed at the immediate vicinity of the NTZ, so mortality by fisheries would be expected to occur as a consequence of spillover or even due to short displacements to the open area. Those species are important fish targets, and S. axillare is one of the main species caught by both spear and hook-and-line fishing in the region (Silveira 2018). S. axillare and L. alexandrei individuals (mostly from scattered patch reefs and aggregated patch reefs) were detected outside the NTZ and had part of their home range and core range beyond its northern border, indicating a potential for fish spillover through those areas (Afonso et al. 2008, La Mesa et al. 2012). Our results thus suggest that



fishing mortality is likely to occur at the unprotected reefs contiguous to the scattered and aggregated patch reef areas.

Emigration to areas outside the detection range of the receivers is also likely related to movements toward the deeper reefs beyond the last line of receivers in the barrier reef area. Snappers in Brazil commonly perform ontogenetic migrations from shallow coastal to deeper reef areas (Frédou & Ferreira 2005, Aschenbrenner & Ferreira 2015). Like many snappers, *L. alexandrei* migrate from estuarine/mangrove areas to coastal reefs (Aschenbrenner et al. 2016a,b), a movement that continues gradually to deeper areas as the species is observed up to depths of 50 m at sizes larger than 33 cm TL (Moura & Lindeman 2007, Fernandes et al. 2012). Therefore, the NTZ in the Tamandaré coral reef complex may act as a stepping stone during the ontogenetic migration. Individuals are likely to remain in the NTZ for a few years, as indicated by the increased densities inside the NTZ, with marked abundance of intermediate sizes (around 20 cm). This results in a buildup of abundance and biomass inside the NTZ, with expected spillover to nearby reefs, but mainly to deeper areas as individuals mature.

Snappers are known to perform reproductive migrations from home sites to aggregation sites during well-defined spawning seasons (Biggs & Nemeth 2016). The length at first sexual maturity for *L. alexandrei* is estimated around 20 cm TL, and the species has a clear spawning season (from November to March, with a peak in February) during which fish may aggregate, as indicated by an increase in fisheries landings for the species in the same period (Fernandes et al. 2012). In the present study, the monitoring period of 6 adult sized L. alexandrei (size range from 21.5 to 25.2 cm TL) overlapped with their spawning season. The loss of detections observed for 2 of those individuals (LU-TALE #01 and #05, both with markedly short weighted residency indices) occurred during the spawning season and could be related to reproductive migrations to aggregation sites outside the NTZ, as suggested for L. apodus in Martinique (Garcia et al. 2014).

For *S. axillare*, the time of residency in the NTZ also seems to be sufficient for a significant reduction in mortality, resulting in an increase in both density and size. The formation of large schools, frequently observed in the single-unit reef area, is also a positive sign of recovery, since schooling behavior may be directly affected by fishing mortality (Guerra et al. 2020). Those schools were formed exclusively by initial-phase females, as shallow reef areas in Tamandaré are occupied mainly by this phase. Terminalphase males are most likely present in the deeper areas of the barrier reef (B. P. Ferreira pers. obs.), where visual surveys were not conducted during this study due to limiting conditions (i.e. low visibility caused by stronger current and wave action). It is thus expected that ontogenetic and reproductive movements to deeper areas are likely to occur.

Several parrotfish species have complex social systems (van Rooij et al. 1996, Mumby & Wabnitz 2002), and movement patterns related to social interactions and reproductive behavior may play an important role in space used by individuals (Afonso et al. 2008). Size at maturation for female *S. axillare* is estimated at 25 cm TL, and reproduction is protracted and

occurs over the entire year (D. Veras unpubl. data). Therefore, most tagged individuals were mature females, and migrations associated with reproduction might be an important factor determining movements and residency times of this species in shallow reef areas. Nine S. axillare were last detected during daylight hours by the receivers on barrier reef or by the receiver located at the easternmost part of the aggregated patch reef area, which could indicate a permanent relocation to the unmonitored deeper reefs inside or outside the NTZ. Also, the lower residency index registered for aggregated patch reef and barrier reef fish might be related to frequent short-term displacements to the deeper portions of the barrier reef area beyond the monitoring ranges of the receivers. Such movements could possibly lead to low survival rates due to spillover movements in deeper areas. Indeed, the unmonitored deeper portion of the barrier reef area contiguous to the NTZ is recognized by local fishermen as a major fishing ground for larger parrotfish (>20 cm).

Acoustic telemetry is recognized as an effective tool for examining fish movements and habitat use at a variety of marine environments, as well as to assess the effectiveness of NTZs for protecting different species (Di Franco et al. 2018). However, acoustic telemetry is still relatively expensive, and the number of available acoustic receivers determines the spatial extent of the monitored area. Moreover, in morphologically complex high-relief habitats such as coral reefs, the detection range of receivers will be affected by the existence of physical barriers to signal propagation (Welsh et al. 2012, Selby et al. 2016). In the present study, the capability to detect crossboundary movements and the extent of fish displacements at unprotected areas might have been limited by the low number of receivers deployed outside the NTZ that resulted in gaps in acoustic coverage, particularly in deeper areas. Similarly, possible relocations towards deeper reefs may have been undetected due to the non-overlapping listening ranges of the last line of receivers in the barrier reef area. Therefore, an expansion of the acoustic array to areas outside the NTZ as well as to deeper reef areas within the NTZ are necessary to better understand the extent of spill-over and cross-shelf movements performed by S. axillare and L. alexandrei, as well as the identification of the pathways utilized during those movements.

Both *S. axillare* and *L. alexandrei* used small home range and core range areas usually corresponding to the fish capture locations. Moreover, although relatively small, reefs within the Tamandaré NTZ were shown to harbor spatially segregated subgroups of individuals for both species. The use of distinct reef areas by groups of fish has been shown for other reef fish species (Egli & Babcock 2004, Hammerschlag-Peyer & Layman 2010, Pillans et al. 2017). Siteattached species may benefit from the continued use of small home range and core range areas by the prompt access to resources (e.g. food and shelter) within a familiar reef area (Kramer & Chapman 1999). Additionally, low overlapping space use among fish subgroups may reduce intraspecific competition and therefore improve the overall fitness of the population (Welsh & Bellwood 2012b).

The mean home range size of *S. axillare* was comparable to the home range estimated for *S. cretense* (La Mesa et al. 2012), but approximately 6 to 30 times greater than previously reported for parrotfish species using acoustic telemetry (Afonso et al. 2008, Welsh & Bellwood 2012b, Garcia et al. 2014, Davis et al. 2017). Mean home range for L. alexandrei was equivalent to that estimated for L. apodus (Garcia et al. 2014) and red snapper L. campechanus (Froehlich et al. 2019), although about 6 to 50 times greater than that observed in other studies on Lutjanus species (Hammerschlag-Peyer & Layman 2010, Hitt et al. 2011a, Topping & Szedlmayer 2011). Such differences in home range sizes might be related to specific environmental and ecological features of the different studied sites and to species-specific requirements and behavior (Zeller 1997). However, it could also be a consequence of the different methods used to detect the acoustic signals and the duration of the monitoring period. A common characteristic of the above-mentioned studies with markedly smaller home range estimates was the utilization of active acoustic tracking instead of a multi-receiver passive monitoring array as used in the present study. During the active tracking, acoustic signals are detected by a hydrophone connected to a receiver on the surface and fish position fixes are recorded using a GPS as fish are individually tracked for short durations (from 1 to several days) (Brownscombe et al. 2019). Therefore, this method identifies fine-scale movement patterns over limited time intervals which may not encompass the entire activity space and consequently provides underestimated home ranges. On the other hand, our home range estimates with passive monitoring acoustic arrays are related to the detection ranges of the receivers and are then subject to a higher level of uncertainty. In the present study, such uncertainties were incorporated into the calculation of the utilization distributions (KUD₉₅ and KUD₅₀) as a kernel bandwidth of 150 m (corresponding to the 50% detection probability range) and, therefore, may have resulted in overestimated home ranges. It is also noteworthy that the implementation of the Vemco Positioning System to obtain fine-scale acoustic telemetry data would highly benefit future work in the region by providing more precise home range estimates.

Low variability in home range and core range sizes was observed among individuals within each species. For *S. axillare*, both home range and core range were not correlated with fish length, as previously observed for S. cretense (Afonso et al. 2008) and steephead parrotfish Chlorurus microrhinos (Welsh & Bellwood 2012b). For adult populations of S. cretense, home range size seemed to be related to individual type of social behavior (i.e. schooling fish or territorial fish) instead of body length (Afonso et al. 2008). On the other hand, positive relationships between fish length and home range size have been reported for snappers such as L. campechanus (Topping & Szedlmayer 2011, Piraino & Szedlmayer 2014, Froehlich et al. 2019) and checkered snapper L. decussatus (Nanami & Yamada 2008), which might be attributed to an increase in resource requirements of larger individuals (Wakeman et al. 1979, Kramer & Chapman 1999). However, other studies on snappers have found no influence of body length on home range size (Hammerschlag-Peyer & Layman 2010, Hitt et al. 2011a), suggesting that other factors including competition (Jones 2005), individual learning and behavior (Parsons et al. 2003, Brown & Laland 2003), and seascape structure (Hitt et al. 2011b, Pittman et al. 2014) likely determine home range variability within populations. The lack of statistical significance in the results reported here are likely to have been influenced by the small number of tagged fish, which is a recurrent issue in acoustic telemetry studies (Luo et al. 2009, Hammerschlag-Peyer & Layman 2010). Moreover, for both species, the minimum fish sizes suitable for tagging was limited by the transmitter sizes in order to not exceed the 2% tag to body mass threshold, and thus tagging was biased toward larger individuals. Further investigation on a higher number of individuals comprising a broader size scale (including juvenile and/or sub-adult individuals present in the NTZ) would provide a better understanding of how home range and core range sizes vary within S. axillare and L. alexandrei populations inhabiting the shallow coastal reefs of Tamandaré.

This study was the first to investigate movements and habitat use of teleost coral reef fishes by means of acoustic telemetry in Brazil and provides invaluable insights on the efficiency and functioning of the NTZ established in the coastal reefs of Tamandaré. The effect of the Tamandaré NTZ in terms of increasing fish density and size was shown; however, our findings also highlight the importance of fisheries management outside the NTZ where fishing pressure is high (Silveira 2018) and capable of promoting a rapid decline in S. axillare and L. alexandrei populations. In addition, due to ontogenetic migrations, protection for areas that shelter older life stages of both species is necessary to maintain healthy stocks capable of continuing to sustain high fishing pressure. Protection of larger areas in order to encompass the reef continuum seems to be necessary, although the implementation of large NTZs or an increase in the size of existing NTZs is not an easy task. Recent work has highlighted the need to protect spawning areas (França et al. 2021) and deeper habitats (Eduardo et al. 2018) on Brazilian coral reef complexes. The establishment of more NTZs along the northeastern coast of Brazil is part of the management plan of the large Coral Coast MPA; this study indicates the importance of considering both isolation and connectivity in the design of new protected areas, as well as the relevance of including a heterogeneity of reef habitats.

Acknowledgements. This work was funded by Conselho Nacional de Desenvolvimento Científico e Tecnológico-Brasil (CNPq) through financing ILTER site 18 (PELD Tamandaré Sustentável): Spatial and temporal dynamics of marine ecosystems: connectivity, resilience and sustainable use in southern Pernambuco (441632/2016-5), and the visiting scholar program (PVE 401594/2014-9). We also thank Projeto Meros do Brasil (supported by Petrobrás S.A. — Programa Petrobrás Ambiental) for the acquisition of some of the acoustic receivers and Texas A&M University at Galveston for the loan of 10 acoustic receivers. We acknowledge the assistance and logistic support during field work provided by the Instituto Recifes Costeiros (Fundação SOS Mata Atlântica e Toyota do Brasil) and Centro de Pesquisa e Conservação da Biodiversidade Marinha do Nordeste (CEPENE/ICMBio). We also thank CNPQ for a PhD grant (140950/2016-8) and postdoctoral fellowship (151219/2021-4) to D.L.L., for a postdoctoral fellowship (PVE 155131/2016-8) to A.L.B.G. and for Productivity Research grant (309216/2018-4) to B.P.F. We thank GEFMAR/DIBIO/ ICMBio for a research grant to S.M.R. This work is part of the Ocean Tracking Network — Brazil.

LITERATURE CITED

- Abecasis D, Erzini K (2008) Site fidelity and movements of gilthead sea bream (*Sparus aurata*) in a coastal lagoon (Ria Formosa, Portugal). Estuar Coast Shelf Sci 79:758–763
- Abecasis D, Bentes L, Erzini K (2009) Home range, residency and movements of *Diplodus sargus* and *Diplodus vulgaris* in a coastal lagoon: connectivity between nursery and adult habitats. Estuar Coast Shelf Sci 85: 525–529
- 🗩 Abecasis D, Bentes L, Lino PG, Santos MN, Erzini K (2013)

Residency, movements and habitat use of adult white seabream (*Diplodus sargus*) between natural and artificial reefs. Estuar Coast Shelf Sci 118:80–85

- Abecasis D, Horta e Costa B, Afonso P, Gonçalves EJ, Erzini K (2015) Early reserve effects linked to small home ranges of a commercial fish, *Diplodus sargus*, Sparidae. Mar Ecol Prog Ser 518:255–266
- Abesamis RA, Russ GR (2005) Density-dependent spillover from a marine reserve: long-term evidence. Ecol Appl 15:1798–1812
- Afonso P, Fontes J, Holland KN, Santos RS (2008) Social status determines behaviour and habitat usage in a temperate parrotfish: implications for marine reserve design. Mar Ecol Prog Ser 359:215–227
- Afonso P, Fontes J, Santos RS (2011) Small marine reserves can offer long term protection to an endangered fish. Biol Conserv 144:2739–2744
- Afonso P, Abecasis D, Santos RS, Fontes J (2016) Contrasting movements and residency of two serranids in a small Macaronesian MPA. Fish Res 177:59–70
- Alós J, March D, Palmer M, Grau A, Morales-Nin B (2011) Spatial and temporal patterns in *Serranus cabrilla* habitat use in the NW Mediterranean revealed by acoustic telemetry. Mar Ecol Prog Ser 427:173–186
- Anderson AB, Bonaldo RM, Barneche DR, Hackradt CW, Félix-Hackradt FC, García-Charton JA, Floeter SR (2014) Recovery of grouper assemblages indicates effectiveness of a marine protected area in Southern Brazil. Mar Ecol Prog Ser 514:207–215
- Anderson AB, Joyeux JC, Floeter SR (2020) Spatiotemporal variations in density and biomass of rocky reef fish in a biogeographic climatic transition zone: trends over 9 years, inside and outside the only nearshore no-take marine-protected area on the southern Brazilian coast. J Fish Biol 97:845–859
- Aschenbrenner A, Ferreira BP (2015) Age, growth and mortality of *Lutjanus alexandrei* in estuarine and coastal waters of the tropical south-western Atlantic. J Appl Ichthyol 31:57–64
- Aschenbrenner A, Ferreira BP, Rooker JR (2016a) Spatial and temporal variability in the otolith chemistry of the Brazilian snapper Lutjanus alexandrei from estuarine and coastal environments. J Fish Biol 89:753–769
- Aschenbrenner A, Hackradt CW, Ferreira BP (2016b) Spatial variation in density and size structure indicate habitat selection throughout life stages of two Southwestern Atlantic snappers. Mar Environ Res 113:49–55
- Barnett LAK, Baskett ML (2015) Marine reserves can enhance ecological resilience. Ecol Lett 18:1301–1310
- Beck H, Feary D, Figueira W, Booth D (2014) Assessing range shifts of tropical reef fishes: a comparison of belt transect and roaming underwater visual census methods. Bull Mar Sci 90:705–721
- Biggs CR, Nemeth RS (2016) Spatial and temporal movement patterns of two snapper species at a multi-species spawning aggregation. Mar Ecol Prog Ser 558:129–142
- Bosiger YJ, McCormick MI (2014) Temporal links in daily activity patterns between coral reef predators and their prey. PLOS ONE 9:e111723
- Brown C, Laland K (2003) Social learning in fishes: a review. Fish Fish 4:280–288
- Brownscombe JW, Lédée EJI, Raby GD, Struthers DP and others (2019) Conducting and interpreting fish telemetry studies: considerations for researchers and resource managers. Rev Fish Biol Fish 29:369–400

- Clarke KR, Gorley RN (2006) PRIMER v6: user manual/tutorial (Plymouth Routines in Multivariate Ecological Research). PRIMER-E, Plymouth
- Dahlgren CP, Eggleston DB (2000) Ecological processes underlying ontogenetic habitat shifts in a coral reef fish. Ecology 81:2227–2240
- Davis K, Carlson PM, Lowe CG, Warner RR, Caselle JE (2017) Parrotfish movement patterns vary with spatiotemporal scale. Mar Ecol Prog Ser 577:149–164
- Di Franco A, Plass-Johnson JG, Di Lorenzo M, Meola B and others (2018) Linking home ranges to protected area size: the case study of the Mediterranean Sea. Biol Conserv 221:175–181
- Di Lorenzo M, Claudet J, Guidetti P (2016) Spillover from marine protected areas to adjacent fisheries has an ecological and a fishery component. J Nat Conserv 32:62–66
- Edgar GJ, Stuart-Smith RD, Willis TJ, Kininmonth S and others (2014) Global conservation outcomes depend on marine protected areas with five key features. Nature 506:216–220
- Eduardo LN, Frédou T, Lira AS, Ferreira BP, Bertrand A, Ménard F, Frédou FL (2018) Identifying key habitat and spatial patterns of fish biodiversity in the tropical Brazilian continental shelf. Cont Shelf Res 166:108–118
- Eggertsen L, Ferreira CEL, Fontoura L, Kautsky N, Gullström M, Berkström C (2017) Seaweed beds support more juvenile reef fish than seagrass beds in a southwestern Atlantic tropical seascape. Estuar Coast Shelf Sci 196:97–108
 - Egli DP, Babcock RC (2004) Ultrasonic tracking reveals multiple behavioural modes of snapper (*Pagrus auratus*) in a temperate no-take marine reserve. J Mar Sci 61:1137–1143
- Feitosa JLL, Ferreira BP (2015) Distribution and feeding patterns of juvenile parrotfish on algal-dominated coral reefs. Mar Ecol 36:462–474
 - Feitoza BM, Rosa RS, Rocha LA (2005) Ecology and zoogeography of deep-reef fishes in Northeastern Brazil. Bull Mar Sci 76:725–742
- Ferguson AM, Harvey ES, Taylor MD, Knott NA (2013) A herbivore knows its patch: Luderick, *Girella tricuspidata*, exhibit strong site fidelity on shallow subtidal reefs in a temperate marine park. PLOS ONE 8:e65838
- Fernandes CAF, de Oliveira PGV, Travassos PEP, Hazin FHV (2012) Reproduction of the Brazilian snapper, *Lutjanus alexandrei* Moura & Lindeman, 2007 (Perciformes: Lutjanidae), off the northern coast of Pernambuco, Brazil. Neotrop Ichthyol 10:587–592
 - Ferreira BP, Maida M (2007) Características e perspectivas para o manejo da pesca na Area de Proteção Ambiental marinha da APA Costa dos Corais. In: MMA/SBF (ed) Áreas aquáticas protegidas como instrumento de gestão pesqueira. Série Áreas Protegidas do Brasil 4. Ministério do Meio Ambiente, Brasília, p 29–39
 - Ferreira BP, Gaspar ALB, Coxey MS, Grillo ACM (2018) Manual de Monitoramento Reef Check Brasil. Ministério do Meio Ambiente, Brasília
- Floeter SR, Halpern BS, Ferreira CEL (2006) Effects of fishing and protection on Brazilian reef fishes. Biol Conserv 128:391–402
- França AR, Olavo G, Rezende SM, Ferreira BP (2021) Spatio-temporal distribution of mutton snapper and dog snapper spawning aggregations in the South-west Atlantic. Aquat Conserv 31:1596–1610
- Frédou T, Ferreira BP (2005) Bathymetric trends of northeastern Brazilian snappers (Pisces, Lutjanidae): implica-

tions for the reef fishery dynamic. Braz Arch Biol Technol 48:787–800

- Frédou T, Ferreira BP, Letourneur Y (2006) A univariate and multivariate study of reef fisheries off northeastern Brazil. J Mar Sci 63:883–896
- Freire KMF, Almeida ZS, Amador JRET, Aragão JA and others (2021) Reconstruction of marine commercial landings for the Brazilian industrial and artisanal fisheries from 1950 to 2015. Front Mar Sci 8:659110
- Froehlich CYM, Garcia A, Kline RJ (2019) Daily movement patterns of red snapper (*Lutjanus campechanus*) on a large artificial reef. Fish Res 209:49–57
- Garcia J, Rousseau Y, Legrand H, Saragoni G, Lenfant P (2014) Movement patterns of fish in a Martinique MPA: implications for marine reserve design. Mar Ecol Prog Ser 513:171–185
- Garcia J, Mourier J, Lenfant P (2015) Spatial behavior of two coral reef fishes within a Caribbean marine protected area. Mar Environ Res 109:41–51
- Gell FR, Roberts CM (2003) Benefits beyond boundaries: the fishery effects of marine reserves. Trends Ecol Evol 18: 448–455
- Gilchrist H, Rocliffe S, Anderson LG, Gough CLA (2020) Reef fish biomass recovery within community-managed no take zones. Ocean Coast Manag 192:105210
- Green AL, Maypa AP, Almany GR, Rhodes KL and others (2015) Larval dispersal and movement patterns of coral reef fishes, and implications for marine reserve network design. Biol Rev Camb Philos Soc 90:1215–1247
- Guerra AS, Kao AB, McCauley DJ, Berdahl AM (2020) Fisheries-induced selection against schooling behaviour in marine fishes. Proc R Soc B 287:20201752
- Halpern BS (2003) The impact of marine reserves: Do reserves work and does reserve size matter? Ecol Appl 13:117–137
- Halpern BS, Warner RR (2003) Matching marine reserve design to reserve objectives. Proc R Soc B 270:1871–1878
- Hamilton RJ, Almany GR, Stevens D, Bode M, Pita J, Peterson NA, Choat JH (2016) Hyperstability masks declines in bumphead parrotfish (*Bolbometopon muricatum*) populations. Coral Reefs 35:751–763
- Hammerschlag-Peyer CM, Layman CA (2010) Intrapopulation variation in habitat use by two abundant coastal fish species. Mar Ecol Prog Ser 415:211–220
- Harasti D, Lee KA, Gallen C, Hughes JM, Stewart J (2015) Movements, home range and site fidelity of snapper (*Chrysophrys auratus*) within a temperate marine protected area. PLOS ONE 10:e0142454
- Harborne AR, Mumby PJ, Kappel CV, Dahlgren CP and others (2008) Reserve effects and natural variation in coral reef communities. J Appl Ecol 45:1010–1018
- Heupel MR, Semmens JM, Hobday AJ (2006) Automated acoustic tracking of aquatic animals: scales, design and deployment of listening station arrays. Mar Freshw Res 57:1–13
- Hitt S, Pittman SJ, Brown KA (2011a) Tracking and mapping sun-synchronous migrations and diel space use patterns of *Haemulon sciurus* and *Lutjanus apodus* in the US Virgin Islands. Environ Biol Fishes 92:525–538
- Hitt S, Pittman SJ, Nemeth RS (2011b) Diel movements of fishes linked to benthic seascape structure in a Caribbean coral reef ecosystem. Mar Ecol Prog Ser 427: 275–291
 - Hobson ES (1975) Feeding patterns among tropical reef fishes. Am Sci 63:382–392

- Honda K, Uy WH, Baslot DI, Pantallano ADS, Nakamura Y, Nakaoka M (2016) Diel habitat-use patterns of commercially important fishes in a marine protected area in the Philippines. Aquat Biol 24:163–174
- Honda K, Uy WH, Baslot DI, Pantallano ADS, Sato M, Nakamura Y, Nakaoka M (2017) Importance of outer reef slopes for commercially important fishes: implications for designing a marine protected area in the Philippines. Fish Sci 83:523–535
- Huijbers CM, Nagelkerken I, Layman CA (2015) Fish movement from nursery bays to coral reefs: a matter of size? Hydrobiologia 750:89–101
- Jones DL, Walter JF, Brooks EN, Serafy JE (2010) Connectivity through ontogeny: fish population linkages among mangrove and coral reef habitats. Mar Ecol Prog Ser 401: 245–258
- Jones KMM (2005) Home range areas and activity centres in six species of Caribbean wrasses (Labridae). J Fish Biol 66:150–166
- Khan JA, Welsh JQ, Bellwood DR (2016) Using passive acoustic telemetry to infer mortality events in adult herbivorous coral reef fishes. Coral Reefs 35:411–420
- Kimirei IA, Nagelkerken I, Trommelen M, Blankers P and others (2013) What drives ontogenetic niche shifts of fishes in coral reef ecosystems? Ecosystems 16:783–796
- Kramer DL, Chapman MR (1999) Implications of fish home range size and relocation for marine reserve function. Environ Biol Fishes 55:65–79
- La Mesa G, Consalvo I, Annunziatellis A, Canese S (2012) Movement patterns of the parrotfish Sparisoma cretense in a Mediterranean marine protected area. Mar Environ Res 82:59–68
- Lea JSE, Humphries NE, von Brandis RG, Clarke CR, Sims DW (2016) Acoustic telemetry and network analysis reveal the space use of multiple reef predators and enhance marine protected area design. Proc R Soc B 283: 20160717
- Lee KA, Huveneers C, Macdonald T, Harcourt RG (2015) Size isn't everything: Movements, home range, and habitat preferences of eastern blue gropers (*Achoerodus viridis*) demonstrate the efficacy of a small marine reserve. Aquat Conserv 25:174–186
- Leleu K, Remy-Zephir B, Grace R, Costello MJ (2012) Mapping habitats in a marine reserve showed how a 30-year trophic cascade altered ecosystem structure. Biol Conserv 155:193–201
- Lester SE, Halpern BS, Grorud-Colvert K, Lubchenco J and others (2009) Biological effects within no-take marine reserves: a global synthesis. Mar Ecol Prog Ser 384: 33–46
- Lippi DL, de Oliveira WDM, da Silveira CBL, Ferreira BP (2021) The use of acoustic telemetry to monitor fish species in marine protected areas: a systematic review with notes on Brazilian perspectives. Arq Ciênc Mar 54: 184–195
- Luo J, Serafy JE, Sponaugle S, Teare PB, Kieckbusch D (2009) Movement of gray snapper *Lutjanus griseus* among subtropical seagrass, mangrove, and coral reef habitats. Mar Ecol Prog Ser 380:255–269
- Malcolm HA, Williams J, Schultz AL, Neilson J and others (2018) Targeted fishes are larger and more abundant in 'no-take' areas in a subtropical marine park. Estuar Coast Shelf Sci 212:118–127
- March D, Alós J, Grau A, Palmer M (2011) Short-term residence and movement patterns of the annular seabream

Diplodus annularis in a temperate marine reserve. Estuar Coast Shelf Sci 92:581–587

- Marshell A, Mills JS, Rhodes KL, McIlwain J (2011) Passive acoustic telemetry reveals highly variable home range and movement patterns among unicornfish within a marine reserve. Coral Reefs 30:631–642
- McClanahan TR, Graham NAJ, Maina J, Chabanet P, Bruggemann JH, Polunin NVC (2007) Influence of instantaneous variation on estimates of coral reef fish populations and communities. Mar Ecol Prog Ser 340:221–234
- Meyer CG, Holland KN, Wetherbee BM, Lowe CG (2000) Movement patterns, habitat utilization, home range size and site fidelity of whitesaddle goatfish, *Parupeneus porphyreus*, in a marine reserve. Environ Biol Fishes 59: 235–242
- ^{*}Meyer CG, Papastamatiou YP, Clark TB (2010) Differential movement patterns and site fidelity among trophic groups of reef fishes in a Hawaiian marine protected area. Mar Biol 157:1499–1511
- Micheli F, Saenz-Arroyo A, Greenley A, Vazquez L, Montes JAE, Rossetto M, Leo GAD (2012) Evidence that marine reserves enhance resilience to climatic impacts. PLOS ONE 7:e40832
- Moura RL, Lindeman KC (2007) A new species of snapper (Perciformes: Lutjanidae) from Brazil, with comments on the distribution of *Lutjanus griseus* and *L. apodus.* Zootaxa 1422:31–43
 - Moura RL, Figueiredo JL, Sazima I (2001) A new parrotfish (Scaridae) from Brazil, and revalidation of *Sparisoma amplum* (Ranzani 1842), *Sparisoma frondosum* (Agassiz, 1831), *Sparisoma axillare* (Steindachner, 1878) and *Scarus trispinosus* Valenciennes, 1840. Bull Mar Sci 68: 505–524
- Mumby PJ, Wabnitz CCC (2002) Spatial patterns of aggression, territory size, and harem size in five sympatric Caribbean parrotfish species. Environ Biol Fishes 63:265–279
- Nanami A, Yamada H (2008) Size and spatial arrangement of home range of checkered snapper *Lutjanus decussatus* (Lutjanidae) in an Okinawan coral reef determined using a portable GPS receiver. Mar Biol 153:1103–1111
- Nanami A, Mitamura H, Sato T, Yamaguchi T and others (2018) Diel variation in home range size and precise returning ability after spawning migration of coral reef grouper *Epinephelus ongus*: implications for effective marine protected area design. Mar Ecol Prog Ser 606: 119–132
- Özgül A, Lök A, Tanrıkul TT, Alós J (2019) Home range and residency of Scorpaena porcus and Scorpaena scrofa in artificial reefs revealed by fine-scale acoustic tracking. Fish Res 210:22–30
- Parsons DM, Babcock RC, Hankin RKS, Willis TJ, Aitken JP, O'Dor RK, Jackson GD (2003) Snapper Pagrus auratus (Sparidae) home range dynamics: acoustic tagging studies in a marine reserve. Mar Ecol Prog Ser 262:253–265
- Pastor J, Verdoit-Jarraya M, Astruch P, Dalias N, Nelva Pasqual JS, Saragoni G, Lenfant P (2009) Acoustic telemetry survey of the dusky grouper (*Epinephelus marginatus*) in the Marine Reserve of Cerbere-Banyuls: informations on the territoriality of this emblematic species. C R Biol 332:732–740
- Pillans RD, Babcock RC, Thomson DP, Haywood MDE, Downie RA, Vanderklift MA, Rochester WA (2017) Habitat effects on home range and schooling behaviour in a herbivorous fish (*Kyphosus bigibbus*) revealed by acoustic tracking. Mar Freshw Res 68:1454–1467

- Piraino MN, Szedlmayer ST (2014) Fine-scale movements and home ranges of red snapper around artificial reefs in the northern Gulf of Mexico. Trans Am Fish Soc 143: 988–998
- Pittman SJ, Monaco ME, Friedlander AM, Legare B and others (2014) Fish with chips: tracking reef fish movements to evaluate size and connectivity of Caribbean marine protected areas. PLOS ONE 9:e96028
- Polunin NVC, Roberts CM (1993) Greater biomass and value of target coral-reef fishes in two small Caribbean marine reserves. Mar Ecol Prog Ser 100:167–176
- * Rhodes KL, McIlwain J, Joseph E, Nemeth RS (2012) Reproductive movement, residency and fisheries vulnerability of brown-marbled grouper, *Epinephelus fuscoguttatus* (Forsskål, 1775). Coral Reefs 31:443–453
 - Ribeiro FP (2004) Composição da biocenose e abundância relativa de peixes capturados com covos nos estados do Rio Grande do Norte e Pernambuco (Brasil). Bol Téc Cient CEPENE 12:113–118 (in Portuguese with English Abstract)
 - Rodgers AR, Kie JG, Wright D, Beyer HL, Carr AP (2015) HRT: Home Range Tools for ArcGIS. Version 2.0. Ontario Ministry of Natural Resources and Forestry, Centre for Northern Forest Ecosystem Research, Thunder Bay
- Rooker JR, Dance MA, Wells RJD, Quigg A and others (2018) Seascape connectivity and the influence of predation risk on the movement of fishes inhabiting a backreef ecosystem. Ecosphere 9:e02200
- *Russ GR, Alcala AC (1996) Marine reserves: rates and patterns of recovery and decline of large predatory fish. Ecol Appl 6:947–961
- Sadovy Y, Domeier ML (2005) Are aggregation-fisheries sustainable? Reef fish fisheries as a case study. Coral Reefs 24:254–262
- Selby TH, Hart KM, Fujisaki I, Smith BJ and others (2016) Can you hear me now? Range-testing a submerged passive acoustic receiver array in a Caribbean coral reef habitat. Ecol Evol 6:4823–4835
 - Silveira MF (2018) Pesca artesanal e manejo: uma abordagem temporal comparativa em Tamandaré—PE. MSc thesis, Federal University of Pernambuco, Recife
- Simpfendorfer CA, Heupel MR, Hueter RE (2002) Estimation of short-term centers of activity from an array of omnidirectional hydrophones and its use in studying animal movements. Can J Fish Aquat Sci 59:23–32
- Steneck RS (1998) Human influences on coastal ecosystems: Does overfishing create trophic cascades? Trends Ecol Evol 13:429–430
- Tickler DM, Carlisle AB, Chapple TK, Curnick DJ, Dale JJ, Schallert RJ, Block BA (2019) Potential detection of

Editorial responsibility: Stephen Wing, Dunedin, New Zealand Reviewed by: 2 anonymous referees illegal fishing by passive acoustic telemetry. Anim Biotelem 7:1

- TinHan T, Erisman B, Aburto-Oropeza O, Weaver A, Vazquez-Arce D, Lowe CG (2014) Residency and seasonal movements in *Lutjanus argentiventris* and *Mycteroperca rosacea* at Los Islotes Reserve, Gulf of California. Mar Ecol Prog Ser 501:191–206
- Topping DT, Szedlmayer ST (2011) Home range and movement patterns of red snapper (*Lutjanus campechanus*) on artificial reefs. Fish Res 112:77–84
- Topping DT, Lowe CG, Caselle JE (2005) Home range and habitat utilization of adult California sheephead, Semicossyphus pulcher (Labridae), in a temperate no-take marine reserve. Mar Biol 147:301–311
- van Rooij JM, Kroon FJ, Videler JJ (1996) The social and mating system of the herbivorous reef fish Sparisoma viride: one-male versus multi-male groups. Environ Biol Fishes 47:353–378
- Villegas-Ríos D, Alós J, March D, Palmer M, Mucientes G, Saborido-Rey F (2013) Home range and diel behavior of the ballan wrasse, *Labrus bergylta*, determined by acoustic telemetry. J Sea Res 80:61–71
- Wakeman JM, Arnold CR, Wohlschlag DE, Rabalais SC (1979) Oxygen consumption, energy expenditure, and growth of the red snapper (*Lutjanus campechanus*). Trans Am Fish Soc 108:288–292
- Welsh JQ, Bellwood DR (2012a) How far do schools of roving herbivores rove? A case study using Scarus rivulatus. Coral Reefs 31:991–1003
- Welsh JQ, Bellwood DR (2012b) Spatial ecology of the steephead parrotfish (*Chlorurus microrhinos*): an evaluation using acoustic telemetry. Coral Reefs 31:55–65
- Welsh JQ, Fox RJ, Webber DM, Bellwood DR (2012) Performance of remote acoustic receivers within a coral reef habitat: implications for array design. Coral Reefs 31: 693–702
- Williams-Grove LJ, Szedlmayer ST (2016) Acoustic positioning and movement patterns of red snapper *Lutjanus campechanus* around artificial reefs in the northern Gulf of Mexico. Mar Ecol Prog Ser 553:233–251
- Wolfe BW, Lowe CG (2015) Movement patterns, habitat use and site fidelity of the white croaker (*Genyonemus lineatus*) in the Palos Verdes Superfund Site, Los Angeles, California. Mar Environ Res 109:69–80
- Worton BJ (1989) Kernel methods for estimating the utilization distribution in home-range studies. Ecology 70: 164–168
- Zeller DC (1997) Home range and activity patterns of the coral trout *Plectropomus leopardus* (Serranidae). Mar Ecol Prog Ser 154:65–77

Submitted: July 28, 2021 Accepted: February 18, 2022 Proofs received from author(s): April 25, 2022